# Molecular Characterization of Resistance Mechanisms Associated With Pleuromutilins Among Gram-Positive Clinical Isolates From the Worldwide SENTRY Surveillance Studies for Lefamulin

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# ABSTRACT

Background: Lefamulin (LEF) is the first pleuromutilin antibiotic under clinical development for IV and oral use in humans. LEF recently completed the first of 2 phase 3 clinical trials for the treatment of community-acquired pacterial pneumonia in adults. This study evaluated the sistance mechanisms associated with elevated LEF MICs in a global collection of surveillance isolates from

Methods: A total of 3.195 staphylococci and 4,489 eptococci were included as part of the LEF Surveillance ogram for 2015–2016. Isolates were tested for sceptibility by CLSI broth microdilution method. A total of 33 isolates with LEF MICs ≥0.5 µg/mL were selected for this study. Isolates had their bacterial genomes sequenced (MiSeg Sequencer, Illumina) and screened in silico for ossible LEF resistance mechanisms.

Results: Many Staphylococcus aureus (7/11; 63.6%) harbored vga(A) (LEF MIC, 0.5-4 µg/mL), while 2 strains carried either vga(E) (LEF MIC, >32 μg/mL) or the enterococcal gene Isa(E) (LEF MIC, 32 µg/mL). One S. aureus had an alteration in L4 (E147K: LEF MIC. 16 μg/mL), whereas 1 isolate (LEF MIC, 0.5 μg/mL) did not have any of the known resistance mechanisms estigated. A total of 8 (57.1%) coagulase-negative vlococci (CoNS) contained vga gene variants (LMU MIC, 1–8 μg/mL). Two S. epidermidis carried the cfr gene alone (LEF MIC, 4 µg/mL) or with multiple mutations in 23S rRNA, L3, and L4 (LEF MIC, 32 μg/mL). Two CoNS did not show any LEF resistance mechanisms (LEF MIC, 0.5 µg/mL), while 2 S. sciuri (LEF MIC, 16-32 µg/mL) carried the intrinsic putative sal(A) efflux pump gene. Many streptococci (10/13; 76.9%) carried Isa(E) (LEF MIC, 2-32 μg/mL), while 1 S. anginosus (LEF MIC, 1 μg/mL) had alterations in both L3 and L4. Two streptococci (LEF MIC. 0.5–0.25 µg/mL) had various alterations at 23S rRNA but outside the LEF binding site.

Conclusions: A small number of isolates (28 of 7.684: 0.36%) from global surveillance studies conducted in 2015–2016 had LEF MIC ≥0.5 μg/mL. The most common sistance mechanisms identified in staphylococci and reptococci were vga and Isa(E), respectively. Importantly no S. aureus isolates harbored cfr. Global surveillance will be conducted to monitor changes over time.

# INTRODUCTION

- Lefamulin is the first pleuromutilin antibiotic for intravenous (IV) and oral use in humans in late-stage clinical development to treat community-acquired bacterial pneumonia (CABP) and acute bacterial skin and skin structure infections (ABSSSI)
- This agent inhibits the bacterial protein synthesis by binding the 50S ribosomal subunit at the A- and P-sites in the peptidyl transferase center (PTC) via an "induced-fit" mechanism, which prohibits the correct positioning of the tRNA as characteristic for pleuromutilin antibiotics
- Lefamulin recently demonstrated noninferiority to moxifloxacin ± linezolid in two phase 3 clinical trials for the treatment of CABP and was safe and well tolerated after IV and oral dosing
- The antibacterial profile of lefamulin covers the most relevant organisms causing CABP, including Gram-positive, fastidious Gram-negative, and atypical respiratory pathogens
- The in vitro activity of lefamulin and comparator agents has been monitored against a global collection of Gram-positive and fastidious Gram-negative organism causing CABP and ABSSSI through the SENTRY Antimicrobial Surveillance Program
- This study evaluated the resistance mechanisms associated with elevated lefamulin minimum inhibitory concentration (MIC) values in a global collection of surveillance isolates from 2015 and 2016

# MATERIALS AND METHODS

#### **Bacterial Isolates**

- A total of 3,195 staphylococci and 4,489 streptococci were included as part of the lefamulin surveillance program for 2015–2016
- 25 Gram-positive isolates from 2015 and 2016 met the MIC screening criteria for molecular characterization with confirmed elevated lefamulin MIC results of ≥1 µg/mL (20 of 7,684; 0.26%; **Table 1**); an additional 5 *Staphylococcus* spp. and 3 viridans group streptococci with a lefamulin MIC value of 0.25–0.5 µg/mL were included
- Bacterial isolate identification was confirmed by standard algorithms supported by matrix-assisted laser desorption ionization—time of flight mass spectrometry (Bruker Daltonics, Bremen, Germany) and genome sequencing

#### **Antimicrobial Susceptibility Testing**

- Isolates were tested for susceptibility by broth microdilution following guidelines in the Clinical and Laboratory Standards Institute (CLSI) M07 (2018) document
- Frozen-form broth microdilution panels were manufactured by JMI Laboratories (North Liberty, IA) and contained cation-adjusted Mueller-Hinton broth (2.5–5% lysed horse blood added for testing streptococci)
- Quality assurance was performed by concurrently testing CLSI-recommended quality control reference strains (Staphylococcus aureus ATCC 29213, Enterococcus faecalis ATCC 29212, and Streptococcus pneumoniae ATCC 49619)

#### **Characterization of Resistance Mechanisms** by Next-Generation Sequencing

- Selected isolates had total genomic DNA extracted by the fully automated Thermo Scientific<sup>™</sup> KingFisher<sup>™</sup> Flex Magnetic Particle Processor (Cleveland, OH, USA), which was used as input material for library construction
- DNA libraries were prepared using the Nextera™ library construction protocol (Illumina, San Diego, CA, USA) following the manufacturer's instructions and were sequenced on a MiSeq Sequencer (JMI Laboratories, North Liberty, IA, USA)
- FASTQ format sequencing files for each sample set were assembled independently using de novo assembler SPAdes 3.9.0, and an in-house designed software was applied to the assembled sequences to align against known macrolide, lincosamide, and streptogramin B (MLS<sub>B</sub>) and pleuromutilin resistance genes
- Additional sequences of intrinsic genes associated with the pleuromutilin binding site, including 23S rRNA (PTC), *rplC* (L3), *rplD* (L4), and *rplV* (L22), were evaluated against a susceptible reference strain of the corresponding species
- All intrinsic 23S rRNA target genes or ribosomal protein amino acid sequences were considered wild type if 100.0% homology with the respective reference sequences was displayed
- Differences were annotated when <100.0% homology</li> was observed

### Multilocus Sequence Typing

- Multilocus sequence typing (MLST) was performed by extracting the previously defined set of 7 housekeeping gene fragments (~500 bp)
- Each fragment was compared to known allelic variants for each locus (housekeeping gene) on the MLST website (PubMLST, https://pubmlst.org)
- An allele sharing 100% genetic identity with a known variant received a numeric designation, and a 7-number sequence (1 for each housekeeping gene) formed an allelic profile, defined as sequence types (STs)
- Isolates containing alleles that did not match an existing sequence in the MLST database were submitted/deposited for allele and ST assignments

# RESULTS

- S. aureus selected for this study (lefamulin MIC, ≥0.5 μg/mL) comprised 11 isolates (0.3% and 0.4%) included in the respective 2015 and 2016 lefamulin surveillance programs, displaying a lefamulin MIC range of 0.5->32 µg/mL (Table 1)
- S. aureus isolates harboring the methyl transferase encoding gene cfr were not observed
- 63.6% (7/11) of *S. aureus* isolates harbored *vga*(A) (lefamulin MIC, 0.5–4 μg/mL), while 2 strains carried either *vga*(E) (lefamulin MIC, >32 μg/mL) or *lsa*(E), which was also identified in Streptococcus agalactiae (lefamulin MIC, 32 μg/mL; Tables 2 and 3)
- One *S. aureus* isolate had an alteration in L4 (E147K; lefamulin MIC, 16 μg/mL), whereas 1 isolate (lefamulin MIC, 0.5 μg/mL) did not have any of the known resistance mechanisms investigated (Tables 2 and 3)

Table 1. Lefamulin MIC Results Obtained During Surveillance Programs for 2015 and 2016

	No. of isolates with lefamulin MIC (μg/mL)																
Organism <sup>a</sup>	0.008	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	>32	Total	MIC <sub>50</sub>	MIC <sub>90</sub>
S. aureus	5	10	745	1,836	300	12	2	3	1	0	0	1		4	2,919	0.06	12
Coagulase-negative staphylococci	2	42	139	68	8	3	3	2	3	3	2	0		1	276	0.03	0.06
S. pneumoniae	5	64	377	1,715	1,513	234	10	5							3,923	0.06	0.12
S. agalactiae	1	21	136	4	3	0	0	0	0	0	2	1			168	0.03	0.03
S. anginosus	0	3	4	5	7	2	3								24		
S. anginosus group	1	1	1	3	3	0	1								10		
S. bovis	2	0	0	0	0	1									3		
S. constellatus	1	2	0	2											5		
S. dysgalactiae	0	3	45	8											56		
S. equinus	0	2													2		
S. gordonii					0	2									2		
S. intermedius	0	1	3	1											5		
S. lutetiensis	1	4	0	0	0	0	1	1							7		
S. mitis group	0	2	1	7	7	9	6								32		
S. mitis/oralis		1	0	1	2										4		
S. parasanguinis	0	2	2	2	1										7		
S. pyogenes	5	80	80												165		
S. salivarius	1	2	3	5	1										12		
S. salivarius group		0	1	8	2										11		
S. salivarius/vestibularis	4	2	2	6	2	1									17		
S. sanguinis				0	2	0	1								3		
S. gallolyticus			0	2	2	2	3	9	13	1	0	0		1	33	1	2

C=minimum inhibitory concentration: MIC<sub>50</sub>=MIC at which 50% of the isolates were inhibited: MIC<sub>90</sub>=MIC at which 90% of the isolates were inhibited. <sup>a</sup>Clinical isolates meeting the MIC inclusion criteria and included in this study are highlighted.

Table 2. MIC Values Obtained for Lefamulin and Comparator Agents Tested Against Isolates Included in This Study

Collection No.	Species	MIC Values (μg/mL)											
		Lefamulina	Clindamycin	Chloramphenicol	Linezolid	Q-D	Retapamulin	Oxacillin	Penicillin	Erythromycin			
975498	S. aureus	0.5 (0.5)	>64	16	1	0.5	≤0.06	>2	>2	>8			
981256	S. aureus	0.5 (0.5)	≤0.5	4	1	0.5	0.5	0.5	2	0.12			
924825	S. aureus	1 (1)	≤0.5	8	1	0.5	2	>2	>2	>8			
953474	S. aureus	1 (1)	≤0.5	4	0.5	0.5	1	≤0.25	≤0.06	>8			
879822	S. aureus	2 (>1)	≤0.5	8	1	0.5	4	≤0.25	≤0.06	0.12			
913640	S. aureus	2 (>1)	>64	8	1	0.5	2	>2	>2	>8			
934242	S. aureus	2 (1)	≤0.5	8	0.5	0.5	2	≤0.25	0.12	0.12			
950457	S. aureus	4 (2)	8	4	1	1	>8	>2	2	0.12			
916083	S. aureus	16 (>1)	>64	8	0.25	1	8	>2	>2	>8			
976441	S. aureus	32 (16)	>64	64	0.5	4	>8	0.5	>2	>8			
972481	S. aureus	>32 (>16)	4	4	1	1	>8	>2	>2	4			
939671	S. cohnii	0.5 (2)	≤0.5	4	1	1	1	1	0.25	>8			
939504	S. epidermidis	0.5 (1)	≤0.5	16	16	≤0.25	0.25	>2	>2	>8			
947675	S. epidermidis	1 (0.5)	16	4	0.5	≤0.25	>8	>2	>2	>8			
951555	S. epidermidis	1 (0.5)	>64	4	0.5	4	1	>2	>2	>8			
955639	S. epidermidis	1 (1)	≤0.5	4	0.5	0.5	1	>2	>2	0.12			
956923	S. epidermidis	2 (0.5)	16	2	0.25	≤0.25	>8	>2	>2	>8			
949426	S. epidermidis	2 (2)	1	4	0.5	≤0.25	>8	≤0.25	0.25	≤0.06			
938399	S. epidermidis	8 (4)	≤0.5	2	0.5	≤0.25	8	>2	>2	>8			
952506	S. epidermidis	8 (4)	1	4	0.5	≤0.25	8	>2	>2	>8			
958510	S. epidermidis	8 (2)	2	4	1	≤0.25	>8	≤0.25	0.25	≤0.06			
934123	S. epidermidis	32 (8)	>64	64	128	1	>8	>2	>2	0.5			
939969	S. haemolyticus	4 (4)	>64	32	2	4	4	>2	>2	>8			
944662	S. sciuri	16 (8)	>64	32	1	0.5	8	>2	>2	>8			
941213	S. sciuri	32 (>16)	≤0.5	4	1	1	>8	>2	>2	0.25			
973516	S. bovis group <sup>b</sup>	0.25 (0.25)	≤0.5	2	1	0.5	0.25	NT	0.06	>32			
965031	S. gallolyticus <sup>c</sup>	32 (>16)	4	4	2	1	>8	NT	0.06	>32			
960742	S. lutetiensis	0.5 (0.5)	≤0.5	2	1	1	0.5	NT	0.06	0.03			
947639	S. anginosus	1 (0.5)	2	4	1	1	0.5	NT	0.03	≤0.015			
982012	S. lutetiensis	2 (1)	1	2	1	0.5	2	NT	0.03	≤0.015			
935554	S. agalactiae	16 (16)	4	2	0.5	0.5	4	NT	0.03	0.03			
935557	S. agalactiae	8 (8)	4	2	1	0.5	4	NT	0.03	0.03			
971459	S. agalactiae	8 (8)	>64	2	1	1	8	NT	0.06	>32			

MIC=minimum inhibitory concentration; MIC<sub>50</sub>=MIC at which 50% of the isolates were inhibited; MIC<sub>90</sub>=MIC at which 90% of the isolates were inhibited; Q-D=quinupristin-dalfopristin; NT=not tested. alnitial lefamulin MIC results obtained during the surveillance studies are within parentheses

bS. bovis group was identified as S. gallolyticus subsp. pasteurianus based on genome sequence S. gallolyticus was identified as S. gallolyticus subsp. pasteurianus based on genome sequence

- A total of 14 coagulase-negative staphylococci (CoNS) with lefamulin MIC values of 0.5—>32 μg/mL were further characterized in this study
- One Staphylococcus cohnii and 1 Staphylococcus epidermidis (lefamulin MIC, 0.5 μg/mL for both) did not show any lefamulin resistance mechanisms; only 23S rRNA alterations (C2534T and G2576T) known to affect the binding of oxazolidinones were observed (Table 3)
- A total of 8 CoNS (57.1%) isolates contained the acquired vga gene variants (lefamulin MIC, 1–8 μg/mL)
- One Staphylococcus haemolyticus isolate carried cfr alone (lefamulin MIC, 4 μg/mL), and 1 S. epidermidis isolate carried the cfr gene with multiple mutations in 23S rRNA, L3, and L4 (lefamulin MIC, 32 µg/mL)
- Two Staphylococcus sciuri isolates (lefamulin MIC, 16–32 μg/mL) carried the intrinsic putative sal(A) efflux pump gene (data not shown)
- Among S. epidermidis isolates, multiple alterations in the 23S rRNA were observed when compared with the wild-type control isolate (Table 4); however, it is unclear whether these polymorphisms affected the susceptibility of pleuromutilin agents and all isolates had concomitant presence of vga or cfr (Table 3)
- Among the streptococci, elevated lefamulin MIC values were only observed within S. agalactiae (3 of 168) and 5 viridans group streptococci, whereas isolates with elevated MIC results were not observed among *S. pneumoniae*, *S. pyogenes*, and the majority of viridans group streptococcal species
- All 3 S. agalactiae and the S. lutetiensis and S. gallolyticus subsp. pasteurianus isolates with lefamulin MIC values of 2–32 μg/mL (Table 3) carried Isa(E) - The S. anginosus isolate (lefamulin MIC of 1 μg/mL) had L3 and L4 alterations as well as multiple alterations in 23S rRNA outside the lefamulin binding site
- The S. bovis group isolate 973516 displaying a lefamulin MIC value of 0.25 μg/mL was later identified as S. gallolyticus subsp. pasteurianus (Tables 2 and 3). The lefamulin MIC of 0.25 µg/mL fits the distribution for S. gallolyticus
- The S. lutetiensis isolate 960742 showed a 23S rRNA at 2359A (Table 3)

Table 3. Molecular Epidemiology and Resistance Mechanism Results for Isolates Included in This Study

Species	Collection No.	Year	MLST <sup>b</sup>	Country	Lefamulin MIC, μg/mL	Re	esistance D	eterminants	Ribosomal Mutations <sup>a</sup>				
						cfr	Isa(E)	vga <sup>c</sup>	23S rRNA	L3	L4	L22	
S. aureus	975498	2016	5	USA	0.5	-	-	-	21G>A, 1557A>T	WT	WT	WT	
S. aureus	981256	2016	4,335	New Zealand	0.5	-	-	vga(A)	21A>G, 1557A>T, 2234A>G	WT	WT	WT	
S. aureus	924825	2015	88	Australia	1	-	-	vga(A)	21A>G, 1557A>T, 2234A>G	WT	WT	WT	
S. aureus	953474	2016	398	France	1	-	-	vga(A)	21A>G, 1557A>T, 2234A>G	WT	WT	WT	
S. aureus	879822	2015	1	Slovenia	2	-	-	vga(A)	21A>G, 2234A>G	WT	WT	WT	
S. aureus	913640	2015	1,148	USA	2	-	-	vga(A)	21A>G, 2234A>G	WT	WT	WT	
S. aureus	934242	2016	1,148	USA	2	-	-	vga(A)	21A>G, 2234A>G	WT	WT	WT	
S. aureus	950457	2016	97	USA	4	-	-	vga(A)	21A>G, 2234A>G	WT	WT	WT	
S. aureus	916083	2015	5	Korea	16	-	-	-	21A>G, 1557A>T, 2234A>G	WT	E147K	WT	
S. aureus	976441	2016	398	Brazil	32	-	+	-	21A>G, 1557A>T, 2234A>G	WT	WT	WT	
S. aureus	972481	2016	398	Germany	>32	-	-	vga(E)	21A>G, 1526A>G, 1557A>T	WT	WT	WT	
S. cohnii	939671	2016	N/A <sup>b</sup>	USA	0.5	-	-	-	C2534T	WT	WT	WT	
S. epidermidis	939504	2016	2	Italy	0.5	-	-	-	G2576T	WT	WT	WT	
S. epidermidis	947675	2016	57	USA	1	-	-	vga(A)	а	WT	WT	WT	
S. epidermidis	951555	2016	87	Czech Republic	1	-	-	vga(A), vga(B)	а	WT	WT	WT	
S. epidermidis	955639	2016	87	Italy	1	-	-	vga(A), vga(B)	а	WT	WT	WT	
S. epidermidis	956923	2016	679	Brazil	2	-	-	vga(A)	а	V188I	WT	WT	
S. epidermidis	949426	2016	255	USA	2	-	-	vga(A)	а	WT	WT	WT	
S. epidermidis	938399	2016	5	USA	8	-	-	vga(A)	а	WT	WT	WT	
S. epidermidis	952506	2016	20	Argentina	8	-	-	vga(A)	а	WT	WT	WT	
S. epidermidis	958510	2016	487	USA	8	-	-	vga(A)	а	WT	WT	WT	
S. epidermidis	934123	2016	5	USA	32	+	-	-	C2534T	H146Q, V154L, A157R	G71 R72ins	WT	
S. haemolyticus	939969	2016	3	Mexico	4	+	-	-	а	WT	WT	A29T	
S. sciuri <sup>d</sup>	944662	2016	N/A <sup>b</sup>	Mexico	16	-	-	-	a	WT	WT	A112E	
S. sciuri <sup>d</sup>	941213	2016	N/A <sup>b</sup>	Australia	32	-	-	-	а	WT	WT	WT	
S. bovis groupe	973516	2016	214	France	0.25	-	-	-	696C>T	WT	WT	WT	
S. gallolyticus <sup>f</sup>	965031	2016	N/A	Spain	32	-	+	-	696C>T	WT	WT	WT	
S. lutetiensis	960742	2016	N/A	Belgium	0.5	-	-	-	224T>C, 2359A>G	WT	WT	WT	
S. anginosus	947639	2016	N/A <sup>b</sup>	USA	1	-	-	-	Multiple alterations <sup>a</sup>	A101E	I148L, S150T	WT	
S. lutetiensis	982012	2016	N/A	Argentina	2	-	+	-	WT	WT	WT	WT	
S. agalactiae	935554	2016	19	Mexico	16	-	+	-	WT	WT	WT	WT	
S. agalactiae	935557	2016	19	Mexico	8	-	+	-	WT	WT	WT	WT	
S. agalactiae	971459	2016	19	Korea	8	-	+	_	WT	WT	WT	WT	

<sup>a</sup>23S rRNA mutational analysis was performed on nucleotide sequences. Additional mutations observed in S. aureus are described in S. aureus are described in S. anginosus were as follows: 150A>G, 152C>A, 164G>T, 434T>G, 957A>T, 1577T>C, 1866A>C, 1867C>G, 1872G>C, 1873T>G, and 2357A>T. Protein sequences were analyzed for annotating L3, L4, and L22.

°Includes vga(A), vga(A)LC, vga(B), vga(C), vga(D), and vga(E). The sal(A) gene was detected in both S. sciuri isolates.

eS. bovis group was identified as S. gallolyticus subsp. pasteurianus <sup>f</sup>S. gallolyticus was identified as S. gallolyticus subsp. pasteurianus

Гable 4. Polymorphi	isms Observed	d Within the 23S rl	RNA Among <i>Staphylococcus</i> spp. Other Than <i>S. aureus</i> Selected for This Study <sup>a</sup>	
Organism	Year	Collection No.	23S rRNA	
S. cohnii	2016	939671	333T>C, 1541T_1542GinsG, 1821A_1822GinsG, 2441C_2442GinsG, 2459T_2460CinsC, 2534T_2535CinsC, 2762G_2763CinsC	
S. epidermidis	2016	934123	669T>C,1236T>C, <b>2534T&gt;C</b>	
S. epidermidis	2016	938399	139C>T, 669T>C, 1236T>C	
S. epidermidis	2016	939504	105G>A, 241G>T,669T>C, 1236T>C, <b>2576G&gt;T</b>	
S. epidermidis	2016	947675	241G>T, 669T>C, 1236T>C	
S. epidermidis	2016	949426	669T>C, 1236T>C	
S. epidermidis	2016	951555	139C>T, 241G>T, 669T>C, 1236T>C	
S. epidermidis	2016	952506	139C>T, 669T>C, 1236T>C, 1638C>T	
S. epidermidis	2016	955639	139C>T, 669T>C, 1236T>C, 2809C>T	
S. epidermidis	2016	956923	669T>C, 1236T>C	
S. epidermidis	2016	958510	241G>T, 669T>C, 1236T>C, 1638C>T	
S. haemolyticus	2016	939969	1486C>T, 2235A>G, 2882T>C	
S. sciuri	2016	941213	WT	
S. sciuri	2016	944662	WT	

<sup>a</sup>Bold selections represent alterations known to affect susceptibility of oxazolidinones

# CONCLUSIONS

- The overall resistance to lefamulin was very low and a small number of isolates (25 of 7,684; 0.33%) from global surveillance studies conducted in 2015–2016 had lefamulin MIC values ≥1 μg/mL
- Lefamulin resistance mechanisms identified in S. aureus isolates included Isa(E), vga(A), vga(E), and alteration in L4 (E147K); vga was the most common determinant observed
- No S. aureus isolates harbored cfr. whereas this was the case for 2 CoNS isolates from USA and Mexico
- Two S. sciuri isolates exhibiting elevated lefamulin MIC values (16–32 µg/mL) did not show any of the resistance mechanisms
- The intrinsic presence of sal(A) may be responsible for the elevated lefamulin MIC results in this species
- Although small in number, results from this study indicate that vga and lsa genes are the most common pleuromutilin resistance mechanisms in staphylococcal and streptococcal clinical isolates, respectively, and global surveillance will be conducted to monitor changes over time

## REFERENCES

- (1) CLSI. M07-A10. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard—tenth edition. Wayne, PA, Clinical and Laboratory Standards Institute, 2015.
- (2) CLSI. M100-S27. Performance standards for antimicrobial susceptibility testing: 27th informational supplement. Wayne, PA, Clinical and Laboratory Standards Institute, 2017.

(3) EUCAST (2017). Breakpoint tables for interpretation of MICs and zone

- diameters. Version 7.0, January 2017. Available at http://www.eucast.org/ clinical breakpoints/. Accessed January 2017. (4) Hot C, Berthet N, Chesneau O (2014). Characterization of sal(A), a novel
- gene responsible for lincosamide and streptogramin A resistance in Staphylococcus sciuri. Antimicrob Agents Chemother. 58: 3335-3341
- (5) Sader HS, Flamm RK (2016). Surveillance of lefamulin activity tested against clinical isolates collected from pediatric and adult patients worldwide (2015). JMI Study Number 17-NAB-02 for 16-NAB-07 study.
- (6) Sader HS, Flamm RK (2017). Surveillance of lefamulin activity tested against clinical isolates collected worldwide (2016). JMI Study Number 16-NAB-01 study. JMI data on file.
- ) Wendlandt S., Lozano C., Kadlec K., Gomez-Sanz E., Zarazaga M., Torres C., Schwarz S (2013). The enterococcal ABC transporter gene Isa(E) confers combined resistance to lincosamides, pleuromutilins and streptogramin A antibiotics in methicillin-susceptible and methicillin-resistant Staphylococcus aureus. J Antimicrob Chemother. 68: 473-475.

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#### **Disclosures**

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